

G-50 Mini Column

For research use only

- Storage** : 2-8°C for up to 6 months. **Do not freeze**
- Sample** : 20 to 50 µl
- Format** : spin column
- Filtrate** : > 20 bases

Geneaid



CERTIFICATE NO. QAIC/TW/50077

www.geneaid.com

Introduction

G-50 Mini Columns consist of prepacked Sephadax G-50 pre-hydrated with double-distilled water. The columns can be used for removing excess dye terminator, free nucleotides from sequencing and labeling reactions, desalting and for buffer exchange. G-50 Mini Columns can purify DNA fragments larger than 20 bases in length with low molecular weight material retained in the gel matrix of the column. Since G-50 Mini Columns are designed to purify DNA fragments >20 bases only, they are not recommended for PCR product primer removal.

Quality Control

The quality of the G-50 Mini Columns is tested on a lot-to-lot basis. The purified DNA is checked by electrophoresis.

Kit Contents

Name	CG002	CG050
G-50 Mini Column	2 pcs	50 pcs
2 ml Collection Tube	2 pcs	50 pcs

Order Information

Product Name	Package size	Cat. No.
G-25 Mini Column	25 preps	CG025
G-50 Mini Column	50 preps	CG050
96-Well G-50 Plate	4/10 x 96-Wells	CGP04/10

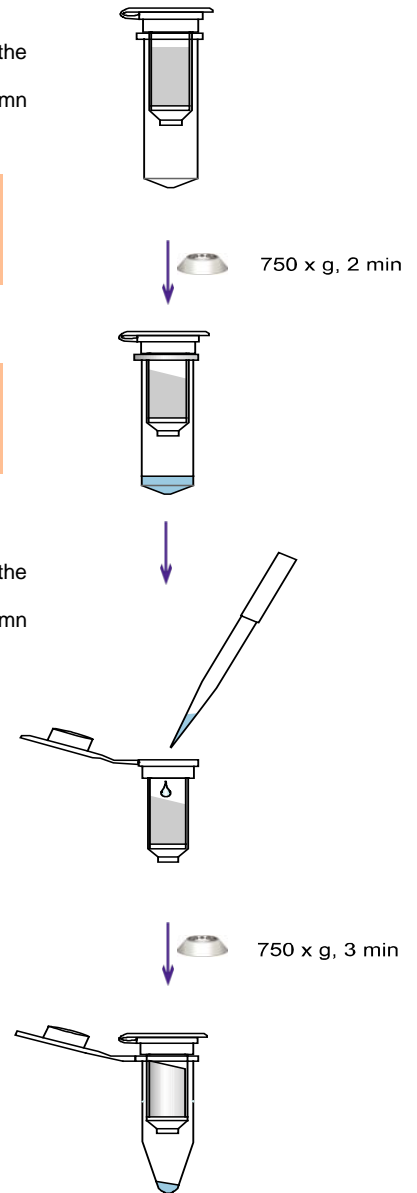
Specifications

During shipment the gel in G-50 Mini Column may be lodged in the cap. This will not affect the quality or the efficiency of the column. In the event of gel drying (cracking), add 300 µl of ddH₂O to the G-50 Mini Column before use. The optimal sample volume is 20 to 50 µl (50 µl maximum).

Purification/ Desalting Protocol

During shipment the gel in G-50 Mini Column may be lodged in the cap. This will not affect the quality or the efficiency of the column. In the event of gel drying (cracking), add 300 µl of ddH₂O to the G-50 Mini Column before use. The optimal sample volume is 20 to 50 µl (50 µl maximum).

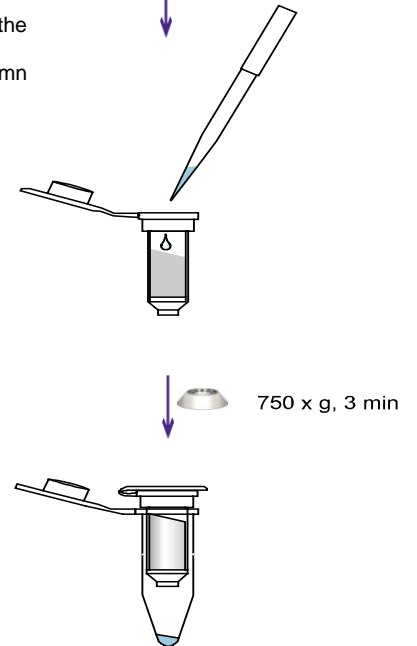
Step 1	<ul style="list-style-type: none"> ● Place a G-50 Mini Column in a 2 ml Collection Tube. ● Centrifuge at approximately 750 x g for 2 minutes (approximately 3000 rpm for an Eppendorf 5415 centrifuge).
Step 2	<ul style="list-style-type: none"> ● Transfer the G-50 Mini Column to a 1.5 ml microcentrifuge tube. ● Carefully load the sample (20-50 µl) onto the center of the gel bed surface.
Step 3	<ul style="list-style-type: none"> ● Centrifuge again at 750 x g for 3 minutes. The purified sample can be recovered at the bottom of the 1.5 ml microcentrifuge tube (approximately the same volume as the loaded sample).



Buffer Exchange Protocol

During shipment the gel in G-50 Mini Column may be lodged in the cap. This will not affect the quality or the efficiency of the column. In the event of gel drying (cracking), add 300 µl of ddH₂O to the G-50 Mini Column before use. The optimal sample volume is 20 to 50 µl (50 µl maximum).

Step 1	<ul style="list-style-type: none"> ● Place a G-50 Mini Column in a 2 ml Collection Tube. ● Centrifuge at approximately 1000 x g for 3 minutes (approximately 3500 rpm for an Eppendorf 5415 centrifuge).
Step 2	<ul style="list-style-type: none"> ● Discard the fraction in the 2 ml Collection Tube and place the G-50 Mini Column back in the same 2 ml Collection Tube.
Step 3	<ul style="list-style-type: none"> ● Add 350 µl of desired buffer to the G-50 Mini Column. ● Centrifuge at approximately 750 x g for 2 minutes.
Step 4	<ul style="list-style-type: none"> ● Transfer the G-50 Mini Column to a 1.5 ml microcentrifuge tube. ● Carefully load the sample (20-50 µl) onto the center of the gel bed surface.
Step 5	<ul style="list-style-type: none"> ● Centrifuge again at 750 x g for 3 minutes. ● The purified sample can be recovered at the bottom of the 1.5 ml microcentrifuge tube (approximately the same volume as the loaded sample).



Troubleshooting

Problem	Possible Reasons/Solution
Gel Drying	<ul style="list-style-type: none"> ● Add 300 µl of ddH₂O to the column before use.